

ALKALOIDS OF *ERYTHROXYLUM CUNEATUM*, *E. ECARINATUM* AND *E. AUSTRALE**

YAHIA M. A. EL-IMAM, WILLIAM C. EVANS and RAYMOND J. GROUT

Department of Pharmacy, The University, Nottingham NG7 2RD, U.K.

(Received 6 October 1987)

Key Word Index—*Erythroxylaceae; Coelocarpus; Erythroxylum cuneatum; E. ecarinatum; E. australe; tropane esters; 3 α ,6 β -dibenzoyloxytropane; 3 α -(4-hydroxyphenylacetoxyl)tropane; 3-cinnamoyloxytropane-6,7-diol; methylecgonidine; nicotine; chemotaxonomy.*

Abstract—From the leaves of *Erythroxylum cuneatum*, a new alkaloid (\pm)-3 α ,6 β -dibenzoyloxytropane was isolated as principal alkaloid; four other benzoyl- and tigloyl-esters were obtained, together with nicotine. Three of these bases were also detected in the stem-bark. Tropacocaine was the principal alkaloid of *E. ecarinatum* leaves. *Erythroxylum australe* root-bark contained the new alkaloids 3 α -(4-hydroxyphenylacetoxyl)tropane and 3-cinnamoyloxytropane-6,7-diol, methylecgonidine as principal base, and other benzoyl- and cinnamoyl-esters.

INTRODUCTION

Section *Coelocarpus* O. E. Schulz [1] of the genus *Erythroxylum* contains some 16 species with a geographical distribution ranging from Australia through SE Asia and India to the eastern side of Africa. Few of the species have received a systematic phytochemical examination although a number have various, including medicinal, native uses [2]. Two such species are *E. cuneatum* (Wall.) Kurz and *E. ecarinatum* Burck. The former is a small tree, widely distributed in SE Asia, with reported uses as a fish poison on Luzon in the Philippines, and as a tonic on the Malay Peninsula [2]. Plowman and Rivier [3] detected neither cocaine nor cinnamoylcocaine in the leaves. *Erythroxylum ecarinatum* is known in Indonesia, New Guinea, the Solomon Islands and Queensland, Australia. The plant has local medicinal and other uses [2] and Australian material gave strongly positive field tests for alkaloids [4]. No detailed examination of the alkaloids of either of these species has been published. On the other hand *E. australe* F. Muell., a small tree of Queensland, has been more extensively studied. In 1889 Maiden [5] recorded that the plant contained no cocaine but that its red wood could be used in cabinet making and as a colourant for cotton and wool. Meteloidine (3a) is the principal alkaloid of the leaves [6] accompanied by smaller amounts of (\pm)-6 β -hydroxy-3 α -tigloyloxytropane (2a), 3 α -hydroxy-6 β -tigloyloxytropane and a base tentatively identified as 7-hydroxy-6-(2-hydroxy-3-phenylpropionyloxy)-3 α -tigloyloxytropane [7]. The root contains 3 α -benzoyloxytropane-6 β ,7 β -diol (3b) [7] and the root-wood various stachene-related diterpenoids [8]. As a further contribution to our studies on the chemotaxonomy and pharmacologically active principles of the genus we now report our findings on the alkaloid composition of various morphological parts of these three species.

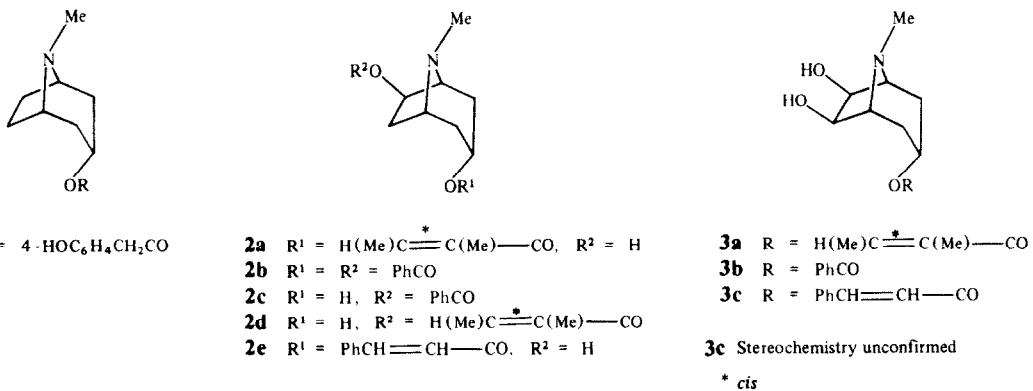
RESULTS AND DISCUSSION

Five bases were characterized from the ether-soluble alkaloid fraction of *Erythroxylum cuneatum* leaves (Table 1). The principal alkaloid was the new natural base (\pm)-3 α ,6 β -dibenzoyloxytropane (2b); its structure was elucidated by spectroscopy and by comparison with the base picrate prepared by partial synthesis. In contrast the corresponding ditigloyl ester found in most *Datura* species is optically active. 6 β -Benzoyloxytropan-3 α -ol (2c) has previously been reported as a constituent of *E. zambeziacum* root-bark [9], *E. glaucum*, *E. cumanense* [10] and *Knightia strobolina*, family Proteaceae [11]. Tropacocaine is a common constituent of the genus but 6 β -tigloyloxytropan-3 α -ol (2d), an alkaloid first isolated from *Datura cornigera* roots [12], has not previously been noted in *Erythroxylum* although its nor-derivative is a constituent of *E. australe* leaves [7]. Nicotine has previously been reported [13] in the young plants, and in roots and stems of mature plants, of *E. coca*; it also co-occurs with tropane alkaloids in section *Salpiglossideae* of the Solanaceae. The present report further demonstrates the apparent sporadic co-occurrence in nature of these two ornithine-derived groups of alkaloids.

The two alkaloids obtained from *E. ecarinatum* leaves (Table 1), tropacocaine and 3 α -benzoyloxytropan-6 β -ol, also co-occur in the leaves of *E. macrocarpum* and *E. sideroxyloides* (section *Pachyllobus*) from Mauritius [14].

Methylecgonidine, the principal alkaloid of the root-bark of *E. australe* (Table 1), was originally isolated from the seeds of *E. coca* [15] and was later reported [16] from *E. dekindtii*. 3 α -Benzoyloxytropane-6 β ,7 β -diol (3b) originally recorded for this species by Griffin [7] has been confirmed for the current sample together with the detection, by mass spectroscopy, of its cinnamoyl analogue (3c), a base not previously known. The molecular ion ($C_{17}H_{21}NO_4$) and the ion at $M - 60$ { $M - [C(6)HOH - C(7)HOH]$ } clearly indicated a tropane-3,6,7-triol, esterified at C-3 by cinnamic acid (acylium ion at m/z 131). The structure of the second new alkaloid 3 α -(4-hydroxyphenylacetoxyl)tropane (1a) was determined by

* Part 8 in the series 'Alkaloids of the Genus *Erythroxylum*'. For part 7 see ref. [9].

Table 1. Alkaloids of *Erythroxylum* sect. *Coelocarpus* O. E. Schulz

	Total alkaloid (% dry weight as tropacocaine)	Alkaloids characterized
<i>Erythroxylum cuneatum</i>		
Leaves	0.08	(±)-3α,6β-Dibenzoyloxytropane (2b) (new alkaloid); 6β-tigloyloxytropan-3α-ol (2d); tropacocaine; nicotine; uncharacterized base involving benzoyl moiety; 6β-benzoyloxytropan-3α-ol (2c)
Stem-bark	0.03	(±)-3α,6β-Dibenzoyloxytropane (2b); tropacocaine; nicotine
<i>E. ecarinatum</i>		
Leaves	0.11	Tropacocaine (principal alkaloid); 3α-benzoyloxytropan-6β-ol
<i>E. australe</i>		
Root-bark	0.30	Methylecgonidine (principal alkaloid); 3α-(4-hydroxyphenylacetoxyl)tropane (1a) (new alkaloid); 3-benzoyloxytropane-6,7-diol (3b); 3-cinnamoyloxytropane-6,7-diol (3c) (new alkaloid); 3α-cinnamoyloxytropan-6β-ol (2e); uncharacterized ester of tropine and acid (C ₉ H ₁₀ O ₃)

spectroscopy. The molecular formula, C₁₆H₂₁NO₃, and the mass fragmentation pattern with ions at *m/z* 140 and 124 indicated a tropan-3-ol esterified with an acid C₈H₈O₃. The ¹H NMR spectrum showed a 3β-proton triplet at δ4.97; the singlet at δ3.51 and well-defined signals for a AA'XX' aromatic proton system at δ 6.75 and 7.11, *J* = 8.5 Hz clearly established the ester as a 4-hydroxyphenylacetate. Supporting evidence for a hydroxyphenylacetate was provided by the ion at *m/z* 107 corresponding to [HOC₆H₄CH₂CO-CO]⁺ [17]. This esterifying acid has not previously been observed in the genus but the corresponding 3-hydroxy-ester is a constituent of *E. hypericifolium* roots [17].

3α-Cinnamoyloxytropan-6β-ol (**2e**) was identified as a minor contaminant in the fraction containing the foregoing ester. By mass spectrometry the molecular formula was established as C₁₇H₂₁NO₃, and the ion at *m/z* 233, representing the loss of the CH₂-CHOH bridge, indicated a derivative of tropane-3,6-diol, with C₉H₈O₂ as the esterifying acid. The ¹H NMR signal at δ5.13 was a triplet, *J* = 5.1 Hz, consistent with a 3α-ester function; the 6β-OH stereochemistry followed from the coupling constants (*J* = 2.5, 7.4 Hz) for the 6α-¹H [17]. The acid

residue gave signals for *trans*-alkene protons at δ6.42 and 7.66, *J* = 16 Hz, and for aromatic protons, in the ratio 2:3, at δ7.4 and 7.55. The (+)-3α,6β-stereoisomer occurs in *Knightia strobolina* (Proteaceae) [11] and as the major alkaloid in the leaves of *E. hypericifolium* [unpublished results]. Also isolated was an unidentified base, *M*, 289, which was an ester of tropan-3-ol. As shown by TLC it was not a tropine ester of tropic acid, 2-hydroxy-3-phenylpropionic acid or 2-, 3- or 4-methoxyphenylacetic acids; the characteristics of the three latter compounds are recorded in the Experimental section. It is of interest that in 1949 Webb reported [4] *E. australe* to exhibit considerable variation in alkaloidal content according to the time and place of collection, a feature confirmed by a comparison of Griffin's study on the roots [7] with that of our own.

It has been suggested [9, 14] that the acids involved in the esterification of the tropanols might prove of intra-generic chemotaxonomic usefulness. In the three species of section *Coelocarpus* studied here benzoic acid appears to predominate and is probably of little chemotaxonomic significance. However the occurrence of tigloyl esters in two species is of interest as they have not so far been

reported in other sections. Further investigations are in progress.

EXPERIMENTAL

Plant material. Leaves and twigs of *E. cuneatum* (Wall.) Kurz collected from a single tree, Lower Bekerong Road, Changi Point, Singapore, Sept. 1980. Air-dried. Voucher: Evans s.n. lodged in City of Nottingham, Natural History Museum, Wollaton. *E. ecarinatum* leaves and twigs from tree, 20–30 m tall, Atherton Tablelands rain forest, 2 km from Yungaburra on road to Malanda, Queensland, Australia, Sept. 1981. Voucher: Plowman and Clarkson No. 10745, lodged in Field Museum of Natural History, Chicago, IL. *E. australis* F. Muell. roots collected Northern Queensland, Australia, 1967. Sample deposited at Pharmacy Department, University of Nottingham, U.K.

Extraction, fractionation and characterization of alkaloids. Ether-soluble alkaloids were extracted from plant material as previously described [10]. Total alkaloids were determined by titration and calculated as tropacocaine. The alkaloid mixtures were resolved by TLC (0.25 mm) and prep. TLC (0.5 mm) as follows: system A: silica gel with $\text{Me}_2\text{CO}-\text{H}_2\text{O}-\text{NH}_4\text{OH}$ (sg 0.88) (80:15:2); system B: Al_2O_3 with $\text{Et}_2\text{O}-\text{EtOH}$ (1:1); system C: Al_2O_3 with Et_2O ; system D: silica gel with $\text{CHCl}_3-\text{Et}_2\text{NH}$ (9:1). Instrumentation and chemical methods were as used previously [17].

E. cuneatum leaves. The leaves (360 g) gave 0.08% total ether-soluble alkaloids resolved into two main fractions by prep. TLC (system D). After further chromatography (system A) the first fraction afforded 6β -tigloyloxytropan-3 α -ol (R_f values in three systems, mp, mmp of picrate, IR); EIMS m/z : 239.1551 (calc. for $\text{C}_{13}\text{H}_{21}\text{NO}_3$; M , 239.1521) and 6β -benzoyloxytropan-3 α -ol (R_f values, mp and mmp of picrate, IR); EIMS m/z : 261.1353 (calc. for $\text{C}_{15}\text{H}_{19}\text{NO}_3$; M , 261.1365). The second main fraction gave four bases (a–d) by system A: (a) uncharacterized R_f 0.3 (system A); IR ν_{max} cm⁻¹: 1715 (ester C=O); EIMS m/z : 122.0985 (calc. for $\text{C}_8\text{H}_{12}\text{N}$; 122.0970), 105.0338 (calc. for $\text{C}_7\text{H}_5\text{O}$; 105.0340); (b) tropacocaine (R_f values, mp, mmp of picrate, IR); EIMS m/z : 245.1423 [$\text{M}]^+$ (calc. for $\text{C}_{15}\text{H}_{19}\text{NO}_2$; M , 245.1416); (c) nicotine [R_f (systems A, B and C) IR]; EIMS m/z : 162.1159 [$\text{M}]^+$ (calc. for $\text{C}_{10}\text{H}_{14}\text{N}_2$; M , 162.1157); (d) characterized as (\pm)-3 α ,6 β -dibenzoyloxytropane; R_f 0.94 (system A); picrate, mp 240° (plates from $\text{EtOH}-\text{H}_2\text{O}$), mmp with synthetic (\pm)-3 α ,6 β -dibenzoyloxytropane picrate (mp 240°) 240° (Found: C, 56.2; H, 4.6; N, 9.4. $\text{C}_{22}\text{H}_{23}\text{NO}_4 \cdot \text{C}_6\text{H}_3\text{N}_3\text{O}$, requires C, 56.6; H, 4.4; N, 9.4%); IR (picrate) ν_{max} cm⁻¹: 1725 (ester C=O); EIMS (probe) 70 eV, m/z (rel. int.): 365.1598 [$\text{M}]^+$ ($\text{C}_{22}\text{H}_{23}\text{NO}_4$ requires M , 365.1627) (7), 229 [picric acid]⁺ (18), 217.1100 (calc. for $\text{C}_{13}\text{H}_{15}\text{NO}_2$; 217.1103) (6), 138 (17), 122.0930 (calc. for $\text{C}_8\text{H}_{12}\text{N}$; 122.0969) (32), 122.0342 (calc. for $\text{C}_7\text{H}_6\text{O}_2$; 122.0368) (9), 105 (66), 94.0643 (calc. for $\text{C}_6\text{H}_8\text{N}$; 94.0656) (100). (\pm)-3 α ,6 β -Dibenzoyloxytropane was prepared from (\pm)-3 α ,6 β -dihydroxytropane and benzoyl chloride, picrate [plates from $\text{EtOH}-\text{H}_2\text{O}$ (1:1)], mp 240° (found: C, 56.5; H, 4.7; N, 9.4%).

E. cuneatum stem-bark. Dried bark (27 g) afforded 0.03% ether-soluble alkaloids. Tropacocaine, nicotine and 3 α ,6 β -dibenzoyloxytropane were identified by TLC (systems A, B and D).

E. ecarinatum leaves. Prep. TLC (system A) of the total ether-soluble alkaloids from 39 g of leaves gave two identifiable bases: (i) tropacocaine (TLC, mp, mmp of picrate, IR); EIMS m/z : 245.1412 [$\text{M}]^+$ (calc. for $\text{C}_{15}\text{H}_{19}\text{NO}_2$; M , 245.1416); (ii) 3 α -benzoyloxytropan-6 β -ol [TLC, IR]; EIMS m/z : 247.1227 [$\text{M}]^+$ (calc. for $\text{C}_{14}\text{H}_{17}\text{NO}_3$; M , 247.1208).

E. australis root-bark. The root-bark (280 g) gave 0.3% ether-soluble alkaloids separable into three groups (X, Y, Z) by prep.

TLC (system B). Rechromatography of X (system A) gave a mixture which was recovered as picrate; it contained 3-benzoyloxytropane-6,7-diol and 3-cinnamoyloxytropane-6,7-diol (R_f 0.71; IR ν_{max} cm⁻¹: 1710 (ester C=O), 3440 (OH)); EIMS (probe) 70 eV, m/z (rel. int.): 303.1456 [$\text{M}_a]^+$ ($\text{C}_{17}\text{H}_{21}\text{NO}_4$ requires M , 303.1471) (4), 277.1324 [$\text{M}_b]^+$ (calc. for $\text{C}_{15}\text{H}_{19}\text{NO}_4$; M , 277.1314) (8), 243.1246 (calc. for $\text{C}_{15}\text{H}_{17}\text{NO}_2$; 243.1259 [M_a – 60]) (4), 229 (picric acid)⁺ (50), 217.1104 (calc. for $\text{C}_{13}\text{H}_{15}\text{NO}_2$; 217.1103 [M_b – 60]) (10), 131.0502 (calc. for $\text{C}_9\text{H}_7\text{O}$; 131.0497) (5), 105.0295 (calc. for $\text{C}_7\text{H}_5\text{O}$; 105.0340) (15), 95.0714 (calc. for $\text{C}_6\text{H}_9\text{N}$; 95.0735) (100), 91 (11). Y gave by rechromatography (system D) two fractions. The first was a mixture of 3 α -(4-hydroxyphenylacetoxyl)tropane, IR ν_{max} cm⁻¹: 1725 (ester C=O), 3420 (OH); EIMS (probe), 70 eV, m/z (rel. int.): 275.1494 [$\text{M}]^+$ ($\text{C}_{16}\text{H}_{21}\text{NO}_3$ requires M , 275.1521) (39), 140.1071 (calc. for $\text{C}_8\text{H}_{14}\text{NO}$; 140.1071) (28), 124.1123 (calc. for $\text{C}_8\text{H}_{14}\text{N}$; 124.1126) (100), 107.0433 (calc. for $\text{C}_7\text{H}_7\text{O}$; 107.0497) [$\text{HOCH}_2\text{CH}_2\text{CO}]^+$ (13); ^1H NMR (250 MHz, CDCl_3), base recovered from picrate: δ 1.58, 1.65, 1.84, 2.2 (4 \times m , H-2, H-4, H-6, H-7), 2.59 (3H, s, NMe), 3.1 (2H, br s, H-5, H-1), 3.51 (2H, s, ArCH_2CO), 4.97 (1H, t, J = 5.2 Hz, H-3 β), 6.75 (2H, d, J = 8.5 Hz, ArH₂), 7.11 (2H, d, J = 8.5 Hz, ArH₂) and a small amount of 3 α -cinnamoyloxytropan-6 β -ol; EIMS (probe) 70 eV, m/z : 287.1514 [$\text{M}]^+$ ($\text{C}_{17}\text{H}_{21}\text{NO}_3$ requires M , 287.1521), 243 [$\text{M}-(\text{CH}_2-\text{CHOH})^+$], 131 [$\text{ArCH}=\text{CHCO}]^+$; ^1H NMR (250 MHz, CDCl_3), base recovered from picrate: δ 2.29 (3H, s, NMe), 2.35 (1H, s, exch. D_2O , OH-6), 3.40 (1H, br s, H-5), 4.64 (1H, dd, $J_{6\alpha}, \gamma_6$ = 2.5 Hz, $J_{6\alpha}, \gamma_2$ = 7.4 Hz, H-6), 5.13 (1H, t, J = 5.1 Hz, H-3 β), 6.42 and 7.66 (2 \times 2H, 2 \times d, J = 16 Hz, *trans* CH = CH), 7.4 (2H, m, m-ArH₂), 7.55 (3H, m, *o*- and *p*-ArH₃), signals for H-2, H-4, H-5 submerged under peaks for similar protons in the foregoing 4-hydroxyphenylacetate. The second fraction of Y gave IR ν_{max} cm⁻¹: 1735 (ester C=O), 3460 (?OH); EIMS (probe) 70 eV, m/z (rel. int.): 289.1688 [$\text{M}]^+$ ($\text{C}_{17}\text{H}_{23}\text{NO}_3$ requires M , 289.1678) (5), 140.1082 (calc. for $\text{C}_8\text{H}_{14}\text{NO}$; 140.1075) (13), 125 (9), 124.1122 (calc. for $\text{C}_8\text{H}_{14}\text{N}$; 124.1126) (100), 96 (17), 94 (19). This base was dissimilar (R_f values) to hyoscyamine and litorine and to the 2-, 3- and 4-methoxyphenylacetates of tropine. Fraction Z gave methylegonidine (TLC, mp, mmp, IR); EIMS m/z : 181.1102 (calc. for $\text{C}_{10}\text{H}_{15}\text{NO}$; M , 181.1103).

Preparation of 2-, 3- and 4-methoxyphenylacetates of tropine. By the method of Hassner and Alexanian [18] esters were prepared by direct esterification of tropine with the relevant acid. The basic products were purified by prep. TLC (system A) and picrates (from EtOH) prepared by standard methods.

3 α -(2-Methoxyphenylacetoxyl)tropane picrate, mp 210° (Found: C, 53.4; H, 4.9; N, 10.5. $\text{C}_{17}\text{H}_{23}\text{NO}_3 \cdot \text{C}_6\text{H}_3\text{N}_3\text{O}_7$ requires C, 53.3; H, 5.05; N, 10.8%). 3 α -(3-Methoxyphenylacetoxyl)tropane picrate, mp 152° (Found C, 53.5; H, 5.1; N, 10.9%). 3 α -(4-Methoxyphenylacetoxyl)tropane picrate, mp 173° (Found: C, 53.2; H, 5.0; N, 10.4%).

Acknowledgements—We are much indebted to the staff of the Singapore Botanical Garden for assisting in locating and collecting the sample of *E. cuneatum*; to Dr T. Plowman (Field Museum, Chicago) for the specimen of *E. ecarinatum* and to Dr J. A. Lamberton (CSIRO, Melbourne) for the supply of the *E. australis* roots.

REFERENCES

1. Schulz, O. E. (1907) in *Das Pflanzenreich* (Engler, A., ed.) Vol. IV, sect. 134. Leipzig.
2. Hagnauer, R. (1981) *J. Ethnopharm.* **3**, 279.
3. Plowman, T. and Rivier, L. (1983) *Ann. Botany (London)* **51**, 641.

4. Webb, L. J. (1949) *CSIRO Aust. Bull.* No. 241, 21.
5. Maiden, J. H. (1889) *The Useful Native Plants of Australia*, p. 176. Turner and Henderson, Sydney.
6. Johns, S. R. and Lamberton, J. A. (1967) *Aust. J. Chem.* **20**, 1301.
7. Griffin, W. J. (1978) *Aust. J. Chem.* **31**, 1161.
8. Connolly, J. D. and Harding, A. E. (1972) *J. Chem. Soc. Perkin Trans. I*, 1996.
9. El-Imam, Y. M. A., Evans, W. C., Grout, R. J. and Ramsey, K. P. A. (1987) *Phytochemistry* **26**, 2385.
10. El-Imam, Y. M. A., Evans, W. C. and Plowman, T. (1985) *Phytochemistry* **24**, 2285.
11. Lounasmaa, M., Pusset, J. and Sévenet, T. (1980) *Phyto-chemistry* **19**, 949.
12. Evans, W. C. and Griffin, W. J. (1963) *J. Chem. Soc.* 4348.
13. Fikenscher, L. N. (1958) *Pharm. Weekblad* **93**, 932.
14. Al-Said, M. S., Evans, W. C. and Grout, R. J. (1986) *Phytochemistry* **25**, 851.
15. Matchett, J. R. and Levine, J. (1941) *J. Chem. Soc.* 2444.
16. Al-Yahya, M. A. I., Evans, W. C. and Grout, R. J. (1979) *J. Chem. Soc. Perkin Trans. I*, 2130.
17. Al-Said, M. S., Evans, W. C. and Grout, R. J. (1986) *J. Chem. Soc. Perkin Trans. I*, 957.
18. Hassner, A. and Alexanian, V. (1978) *Tetrahedron Letters* 4475.